

# Matrix effects in practice: sense and nonsense of internal standard response acceptance criteria

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# Introduction

- The internal standard (IS) is used to compensate for any variability during sample preparation, injection and detection.
- IS is selected to mimic the analyte as closely as possible.
- If IS closely tracks the analyte, why monitor IS response?
  - Inconsistent IS response points to issues with the assay
  - Impact on reliability of the results?
  - IS will not compensate for human error

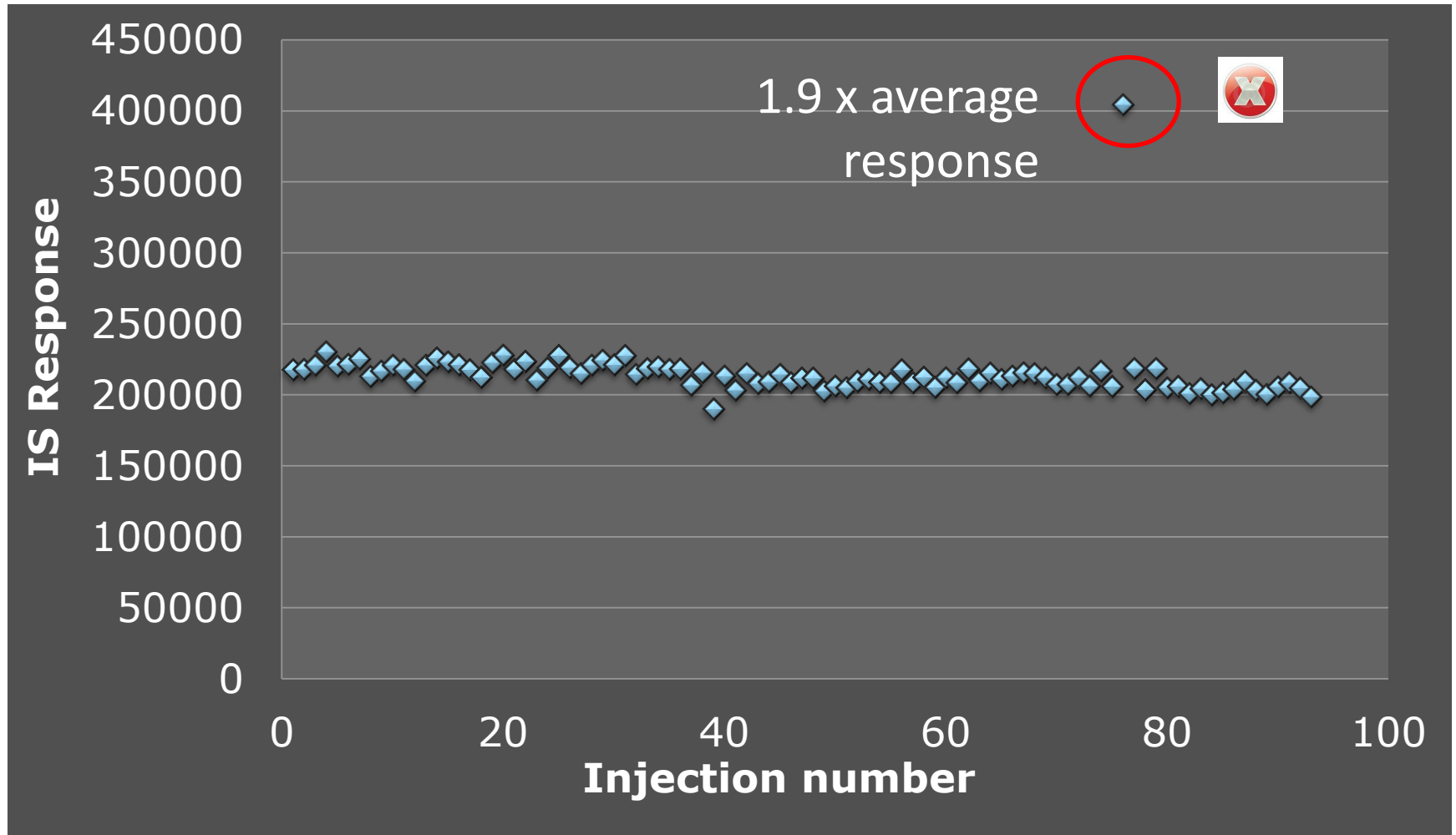
# What do guidances say?

- Workshop/Conference Report on the 3<sup>rd</sup> AAPS/FDA Bioanalytical Workshop (*C.T. Viswanathan et al. The AAPS Journal 2007; 9 (1) Article 4* (<http://www.aapsj.org>))
  - Mentioned under “Validation topics with no consensus” is Acceptance criteria for internal standards:
    - ...in case of low IS response it is important to determine that the assay continues to have the ability to accurately quantify the LLOQ...
    - ...If study samples or analytical runs are rejected or repeated based on IS response variability, objective criteria are necessary and need to be established a priori...
- Although there was no consensus on the topic, it is good practice to have some criteria in place to exclude subjectivity in accepting/rejecting results

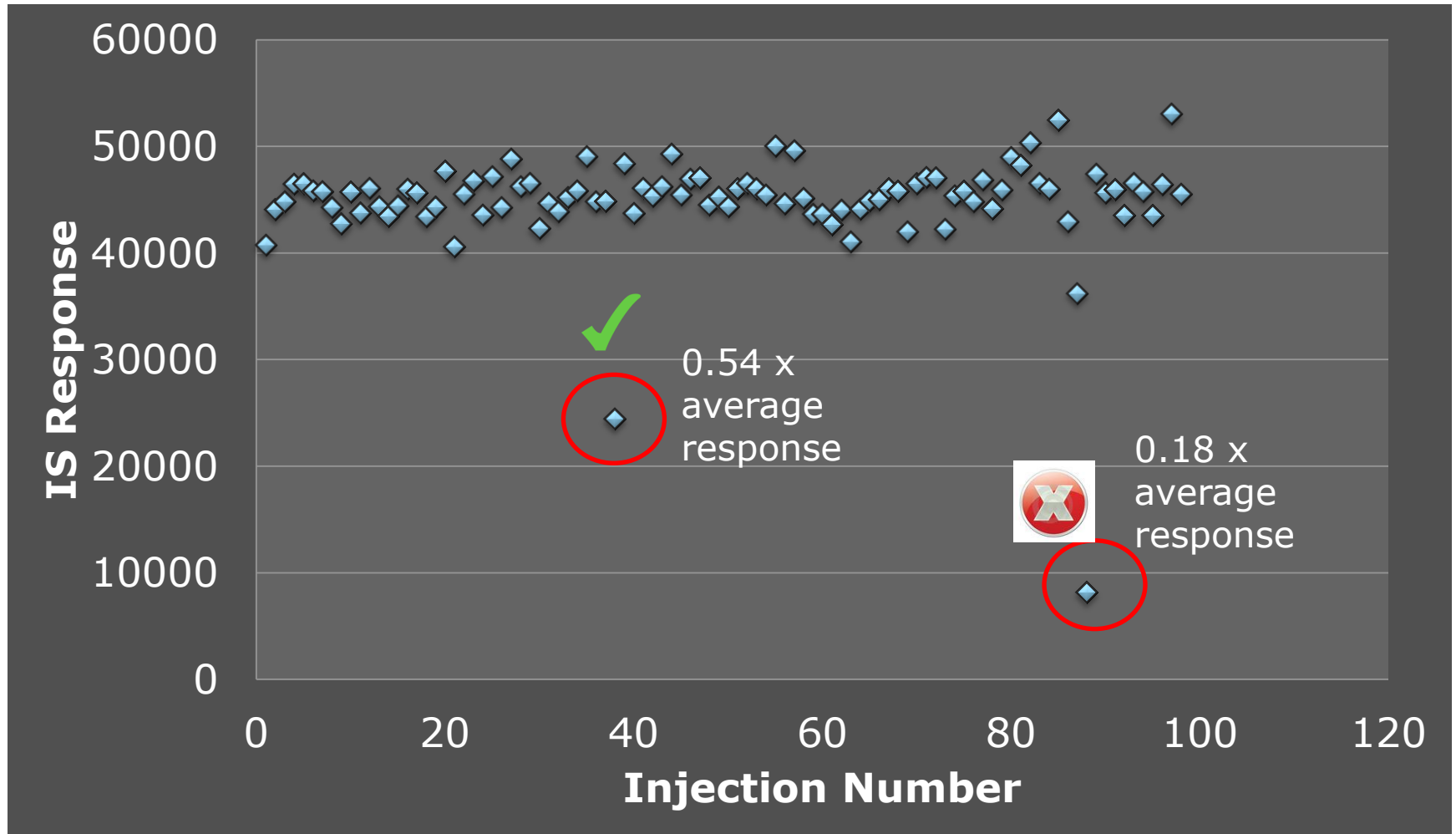
# IS acceptance criteria currently used by Janssen Research and Development

- Criteria straightforward and easy to use; no complicated calculations
- Criteria should enable to filter out gross errors (e.g. spiking errors)
  - Calculate average IS response over the entire batch
  - Lower limit: if IS response  $< 25\%$  (STIL) or  $50\%$  (analog) of average then reject; ability to accurately quantify the LLOQ
  - Upper limit: if IS response  $> 175\%$  of average then reject
  - Do not reject calibration standards or QCs based on this criterion; BCV will tell you if IS compensates for response variations in these samples

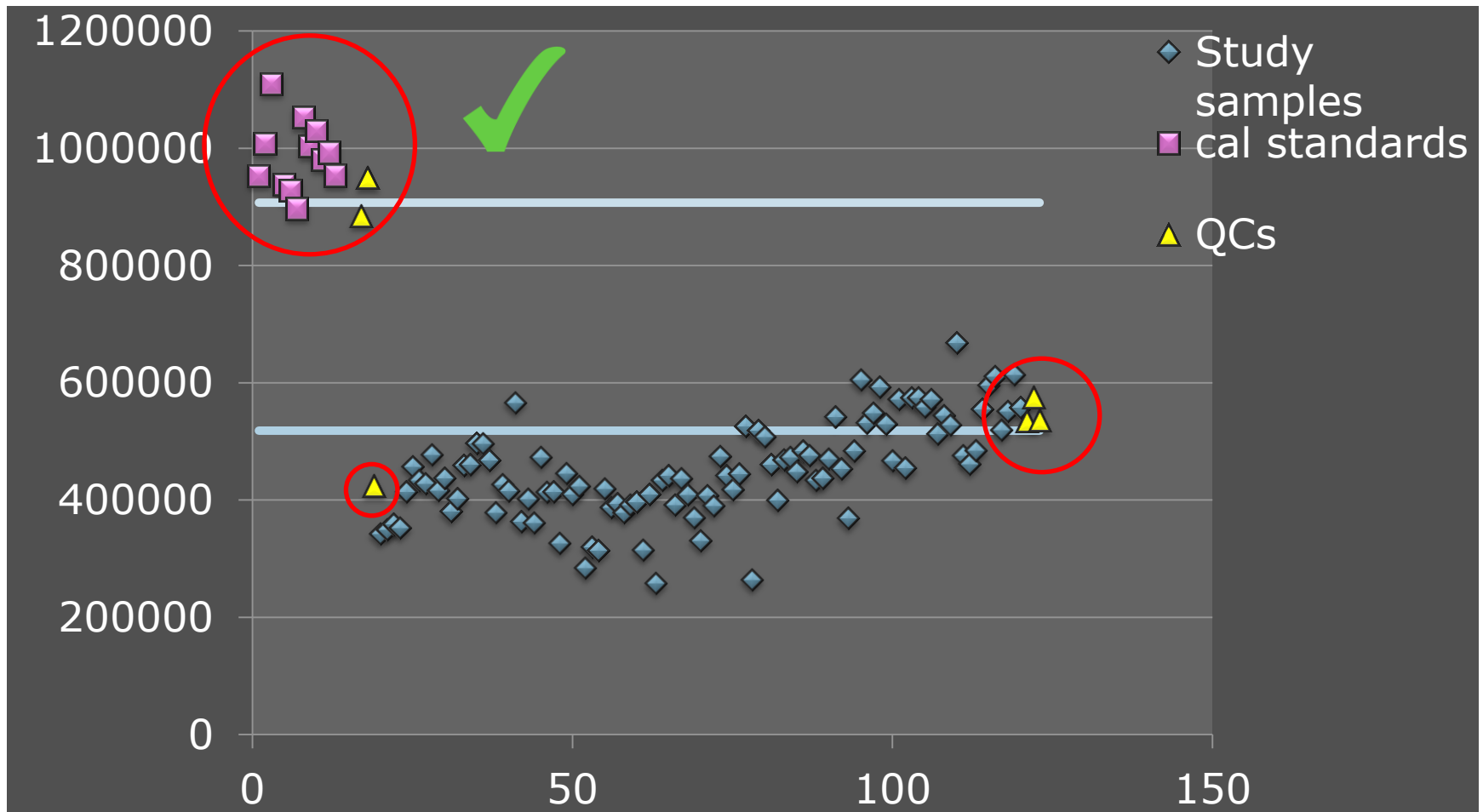
# Example 1



# Example 2



# Example 3

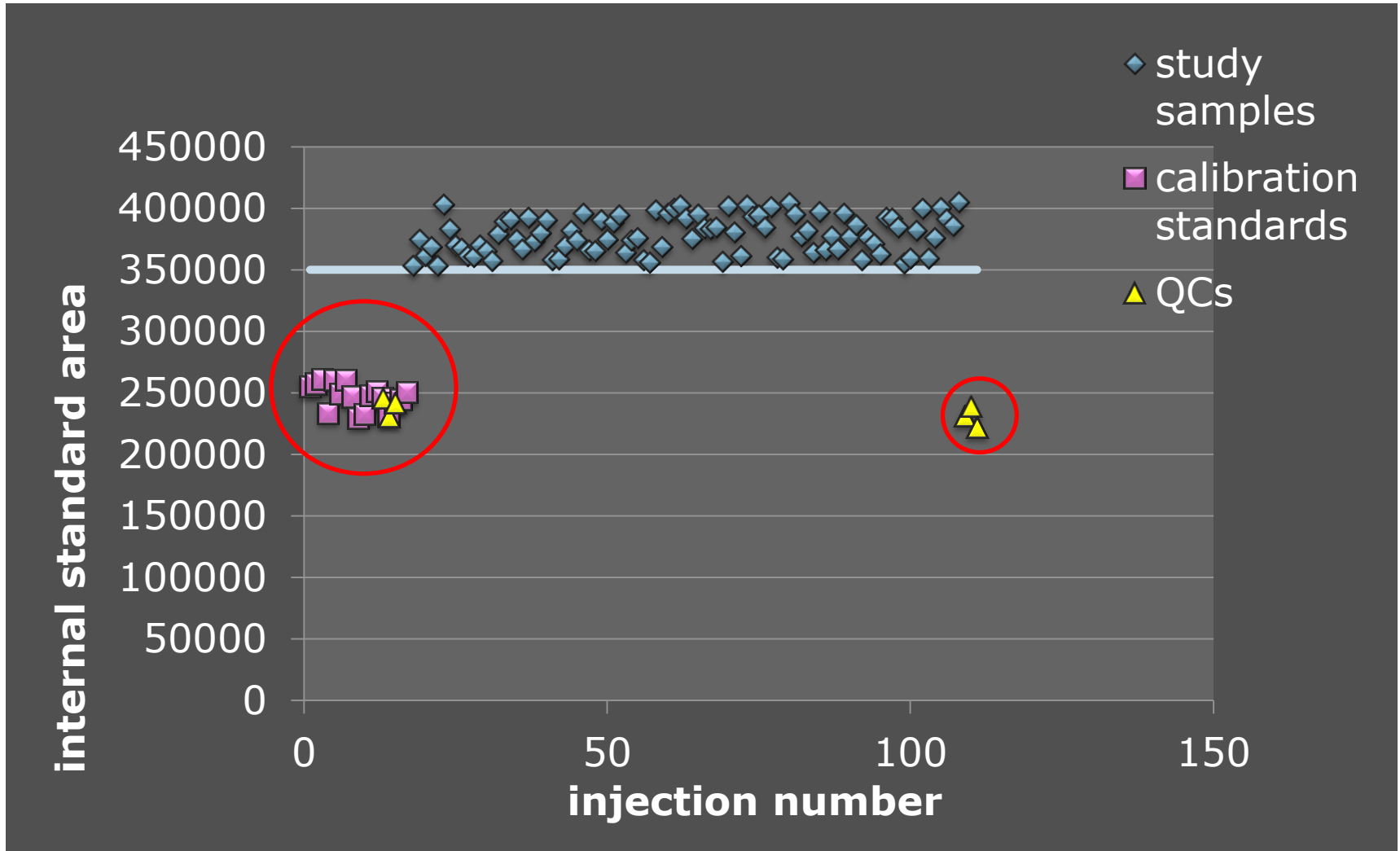




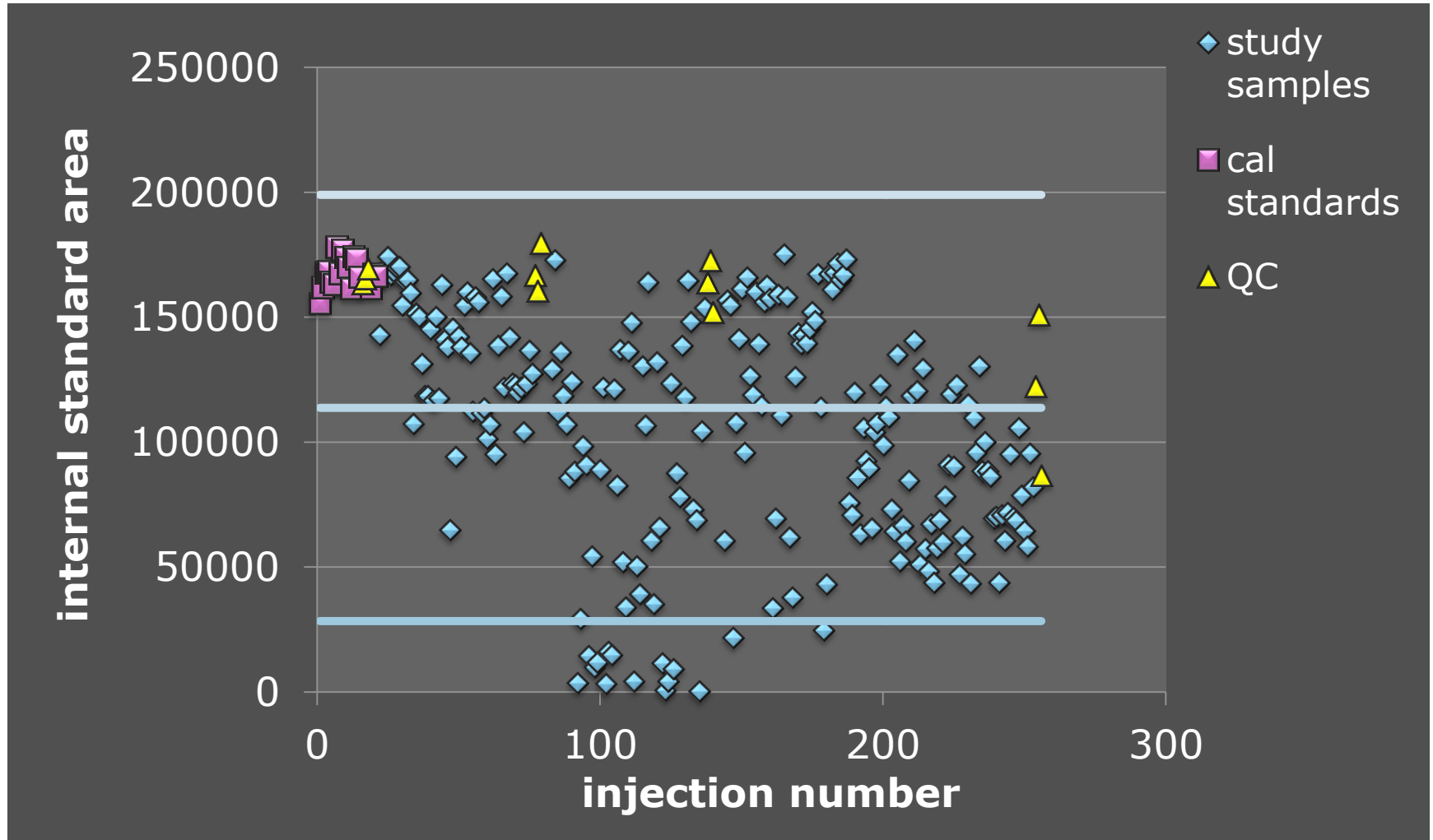
# Are these criteria ideal?

- No, need some fine tuning
- Even when acceptance criteria are met, IS response differences can question reliability of the results
- Additional criteria needed to flag batches where further investigation into root cause is required
- Outcome of root cause investigation could be to
  - accept batch
  - re-inject (instrument problem)
  - re-analyse
- If problem cannot be solved method may require re-development

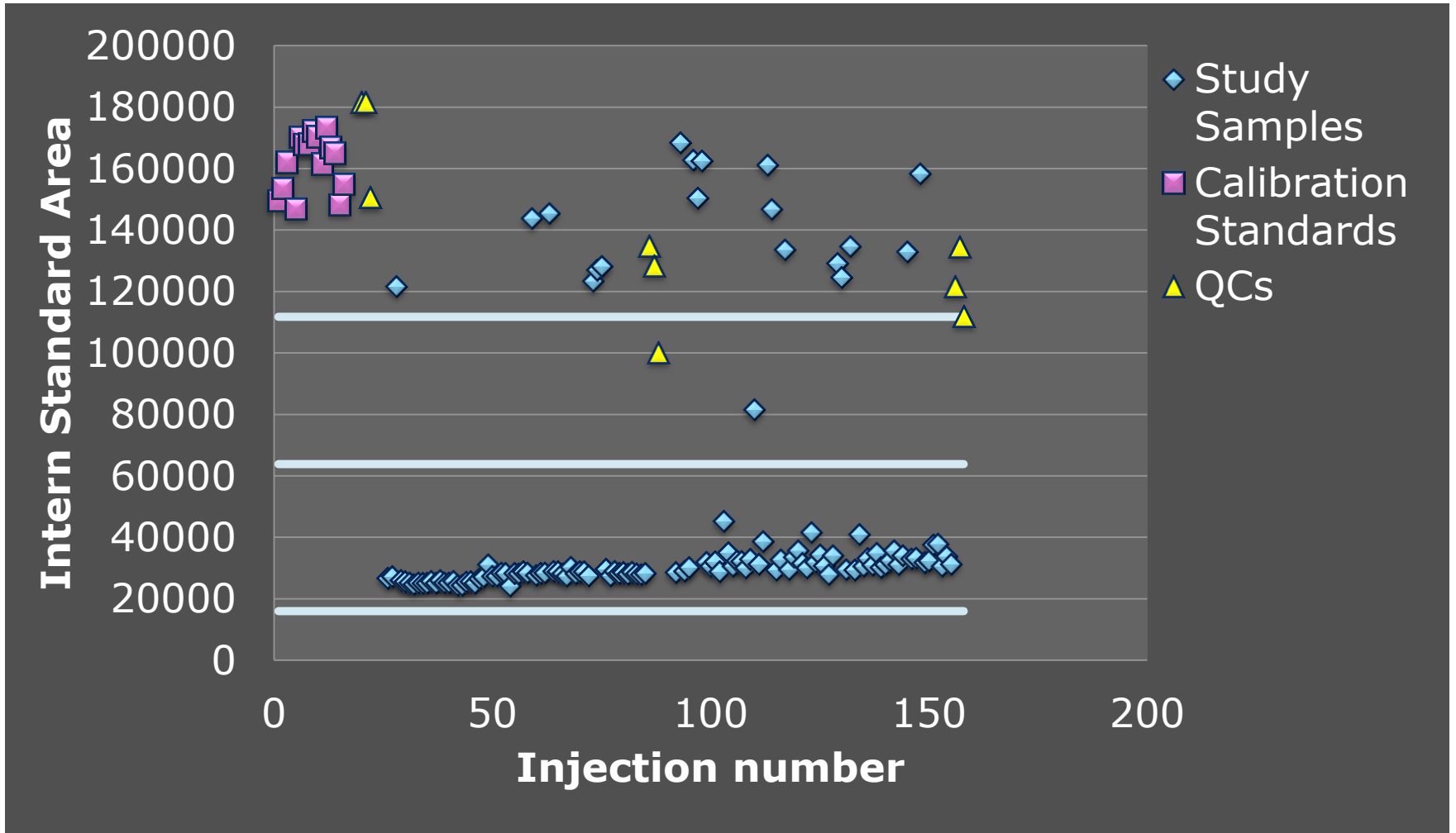
# Example 4



# Example 5



# Example 6



# Fine tuning of criteria

- Criteria should pick up
  - Difference in IS response standards-QCs-study samples
  - Difference in IS response between subjects
  - Large variability in IS response overall
  - Gradient in IS response throughout the run
- Add criteria to initiate further investigation (no hard criterion to reject the batch):
  - 1. overall %CV < 30%
  - 2. average response study samples within 70 – 130% average response standards & QCs
- If one of the above is not met: investigate
- If both of the above are met, apply lower & Upper IS response criterion to accept/reject study samples

# Conclusion

- Having a priori IS response criteria adds value
- Criteria currently used at Janssen R&D are being fine tuned
  - Start by applying first set of criteria to flag unexpected IS behavior requiring further investigation
  - If first criteria pass, apply second set of criteria to reject individual samples based on lower and upper limits
- Proposed criteria should be easy to apply